

# CMOS electronics probe inside biological cellular networks (1<sup>st</sup> generation device)

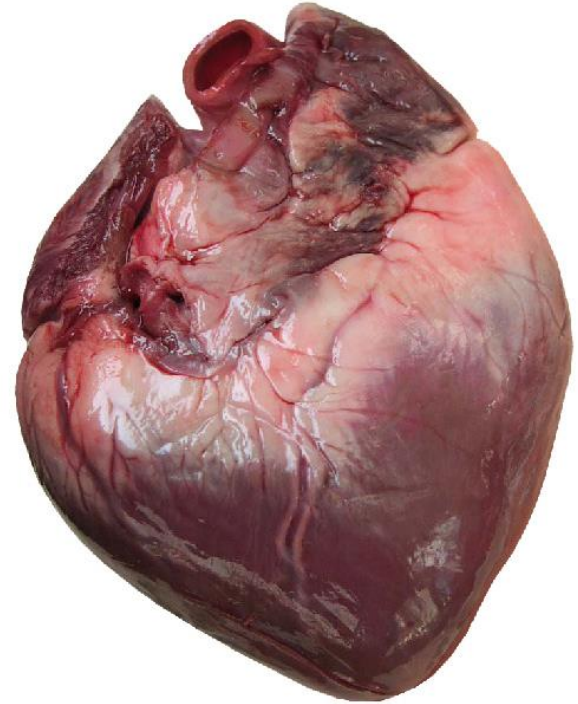
Donhee Ham, Harvard University



# Electrogenic cellular networks



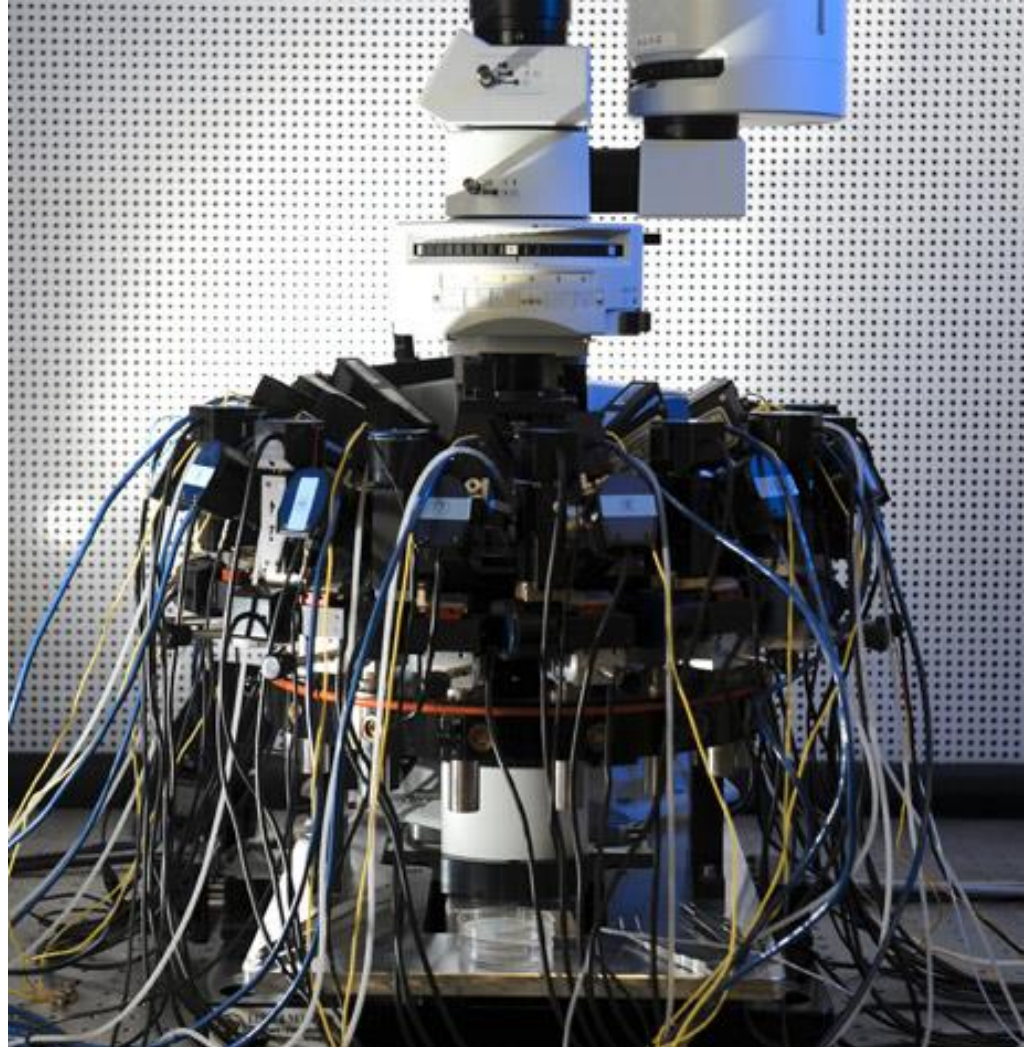
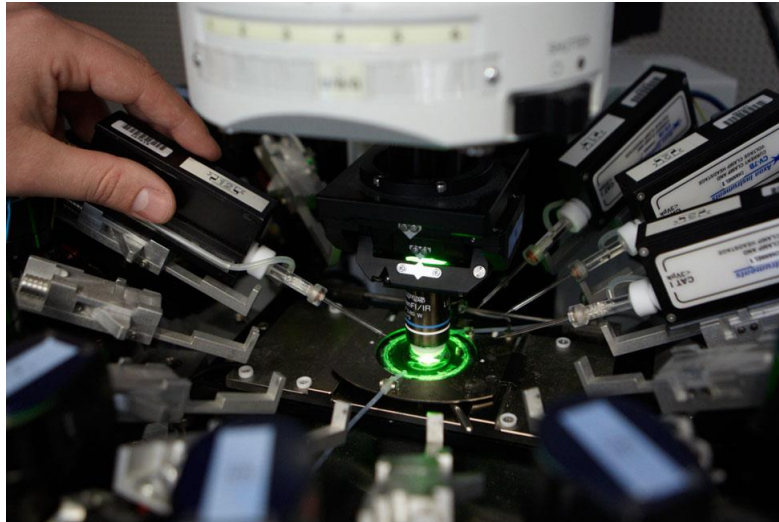
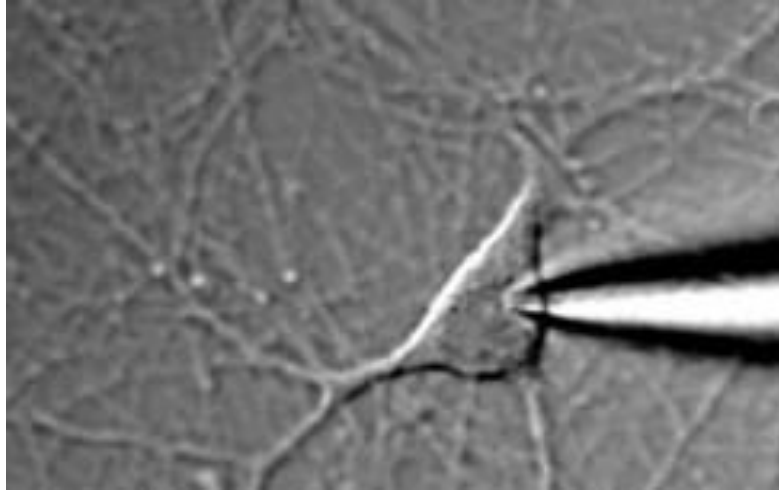
$\sim 10^{11}$  neurons  
 $\sim 10^{15}$  synapses



$\sim 10^9$  cardiomyocytes  
 $\sim 10^{10}$  cell-cell connections

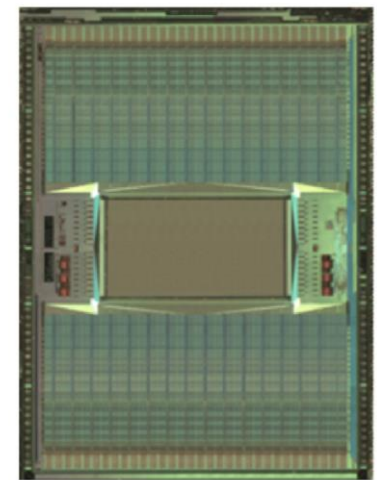
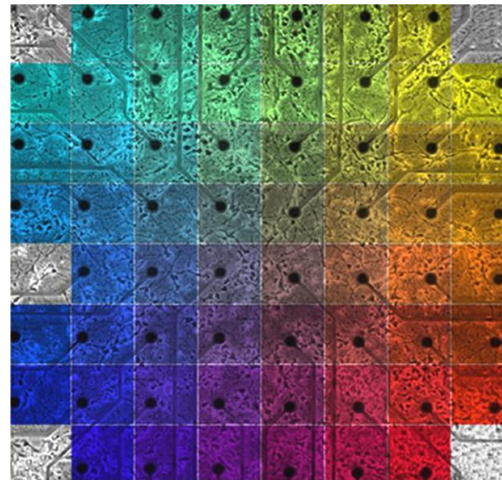
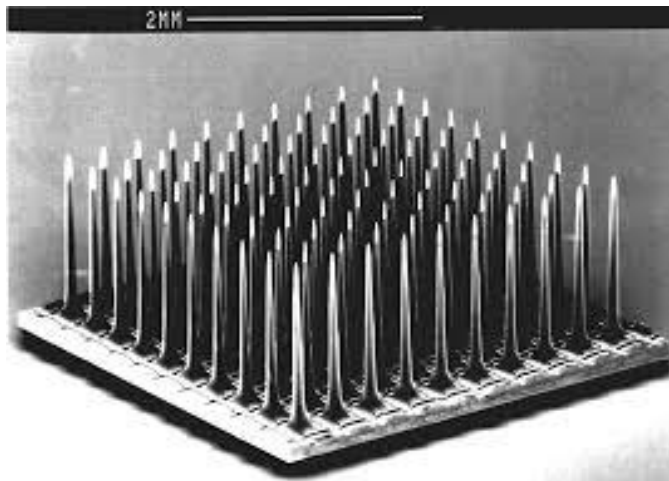
Dichotomy — intracellular vs. parallel

Patch pipette — Intracellular, but not parallel



Dichotomy — intracellular vs. parallel

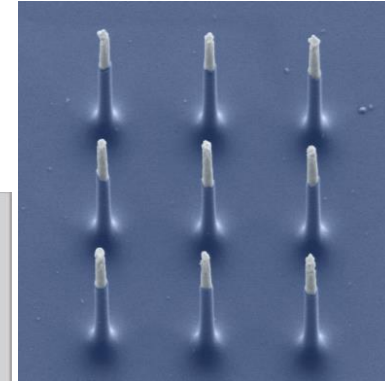
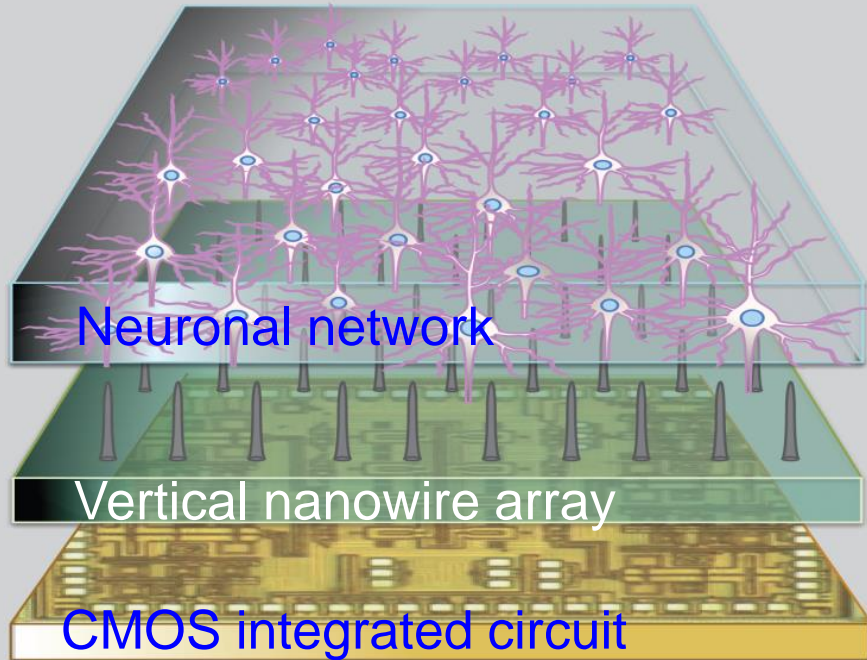
Microelectrode array — parallel, but not intracellular



26,400 electrodes  
1,024 channels

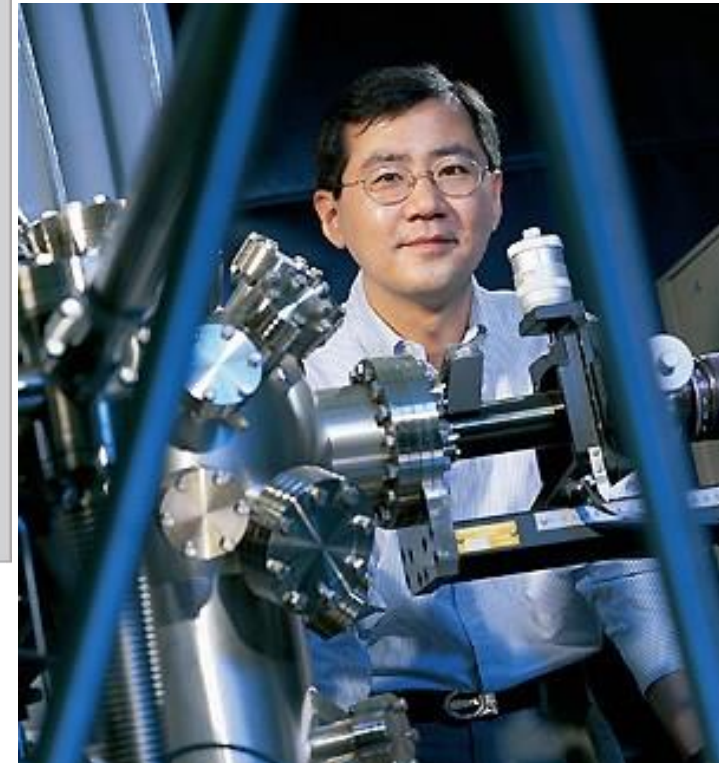
M. Ballini et al.,  
*IEEE JSSC* (2014)

# CMOS nanoelectrode array — Intracellular + parallel



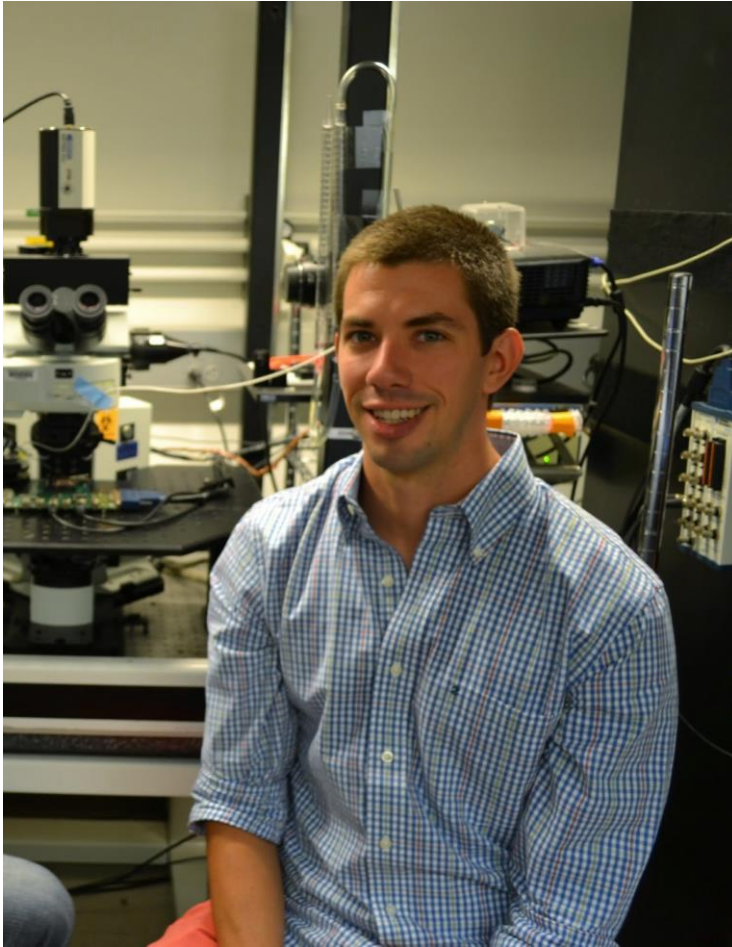
Vertical  
Nanowires

Park lab,  
*Nature Nano.*  
7, 180 (2012).

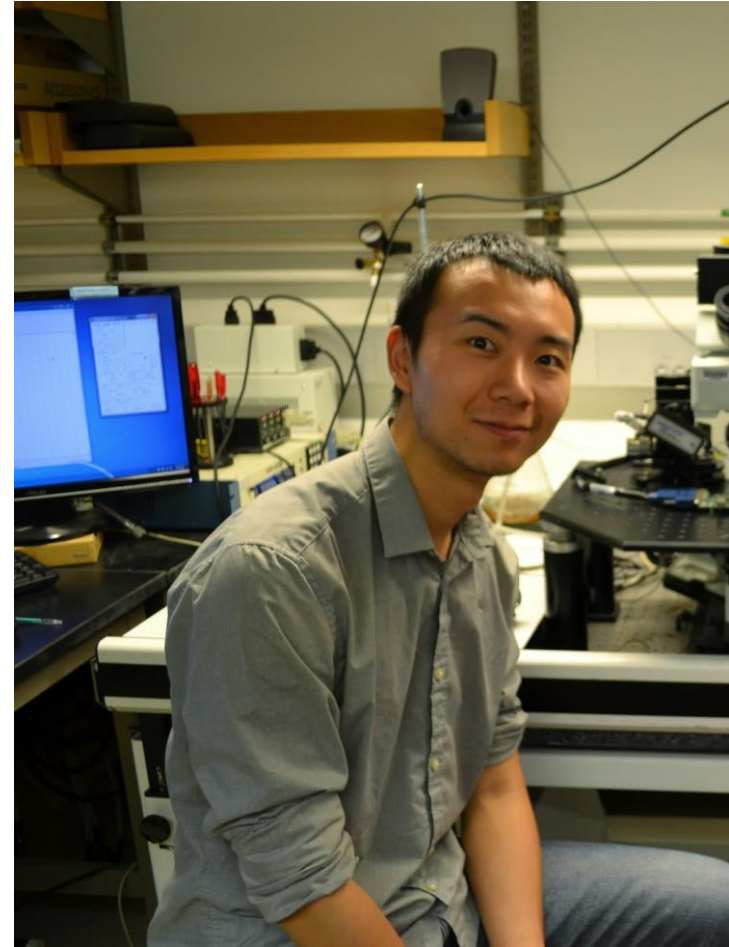


Prof. Hongkun Park  
(Harvard Chemistry & Physics)

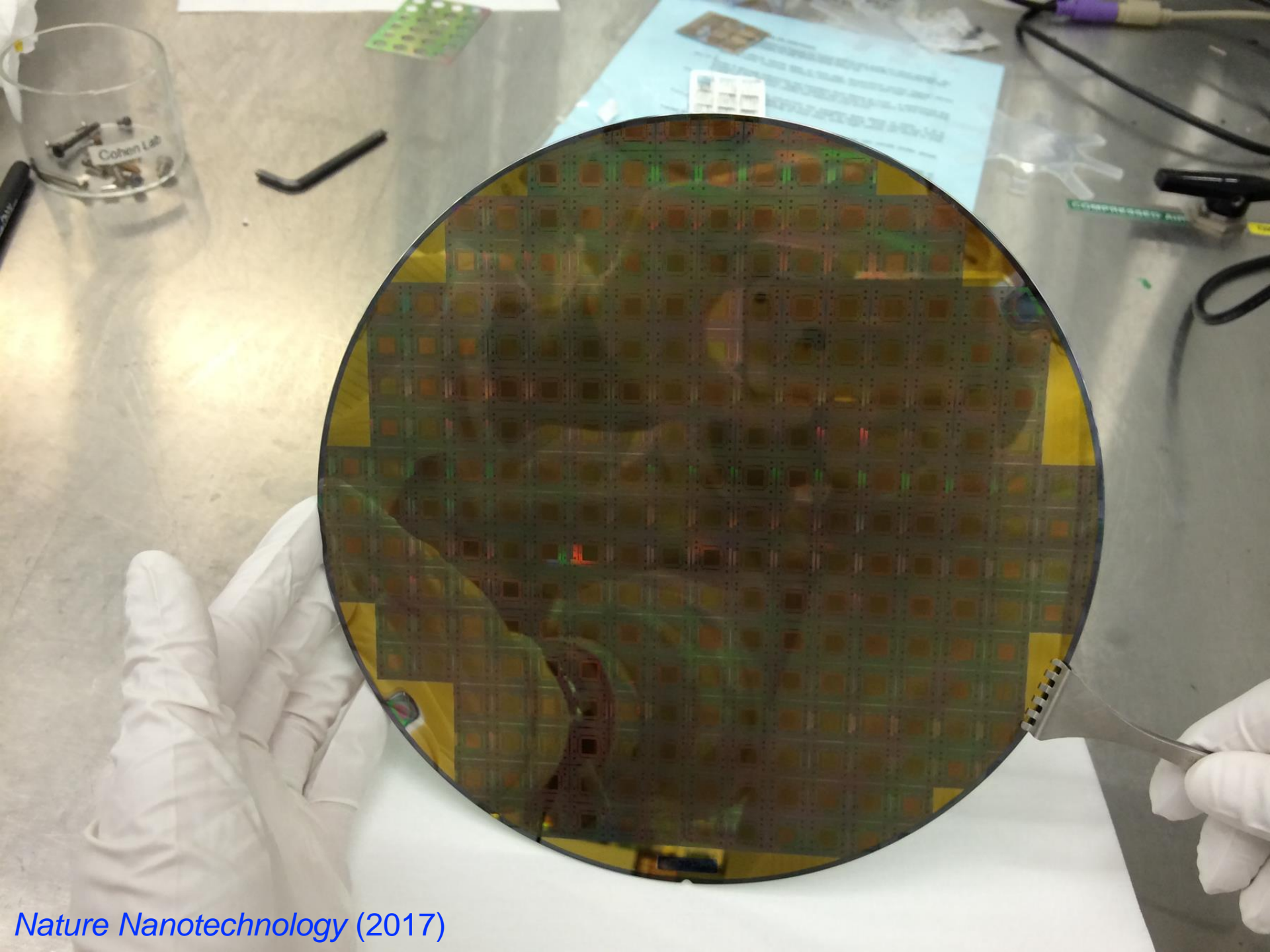
# CMOS nanoelectrode array — Intracellular + parallel



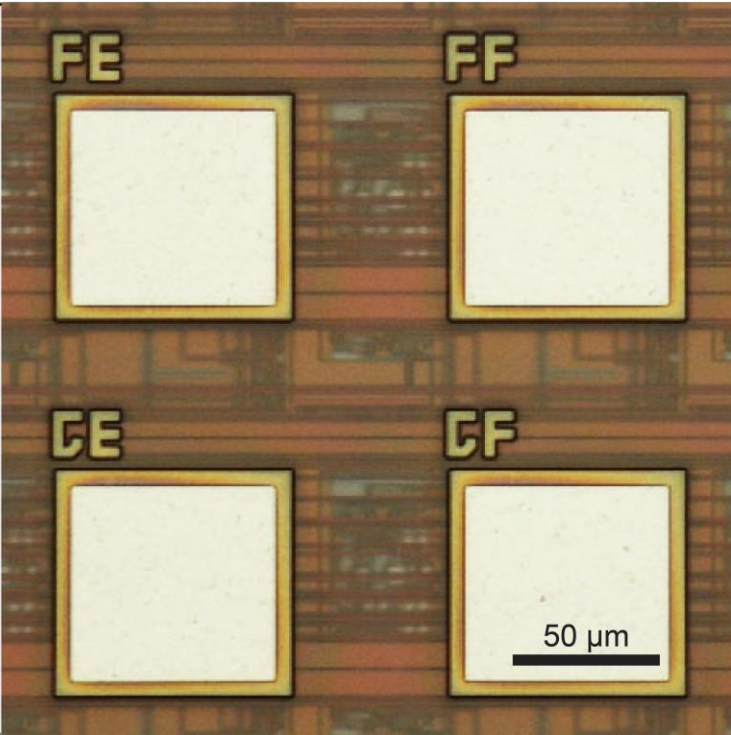
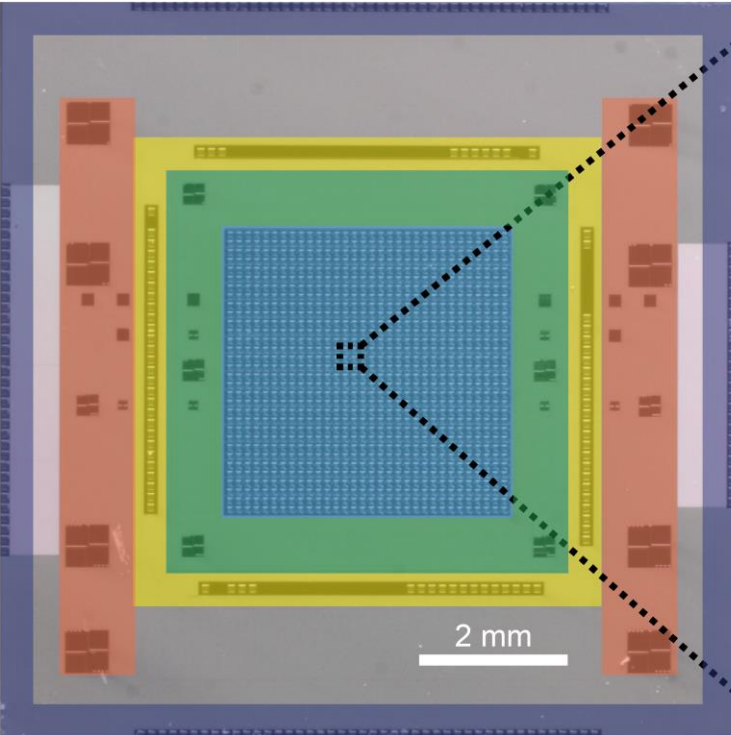
Jeffrey Abbott



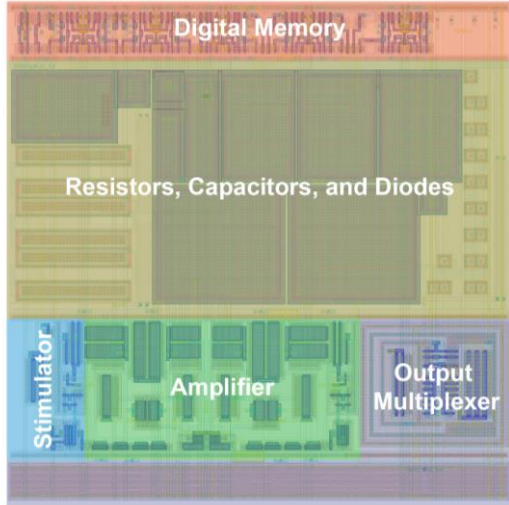
Tianyang Ye (Park lab)



# CMOS IC chip (1024 active site array)

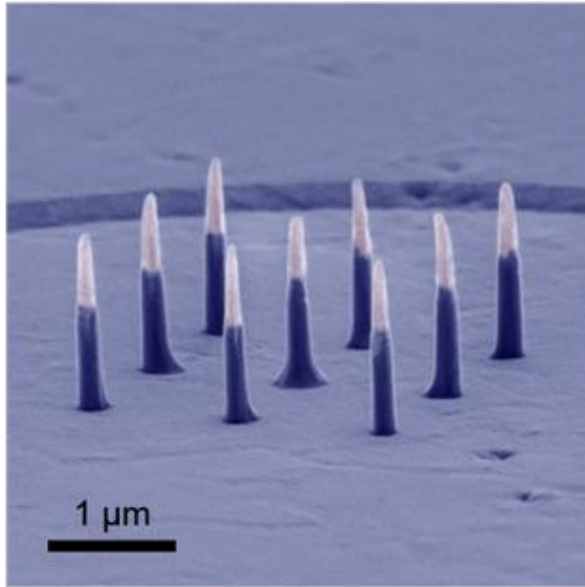


- Array of pixels
- Control Circuitry
- I/O Pads
- Alignment Marks
- Test Structures

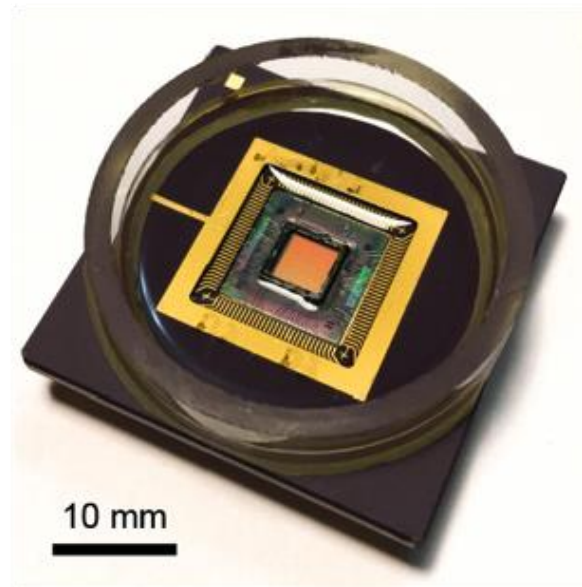




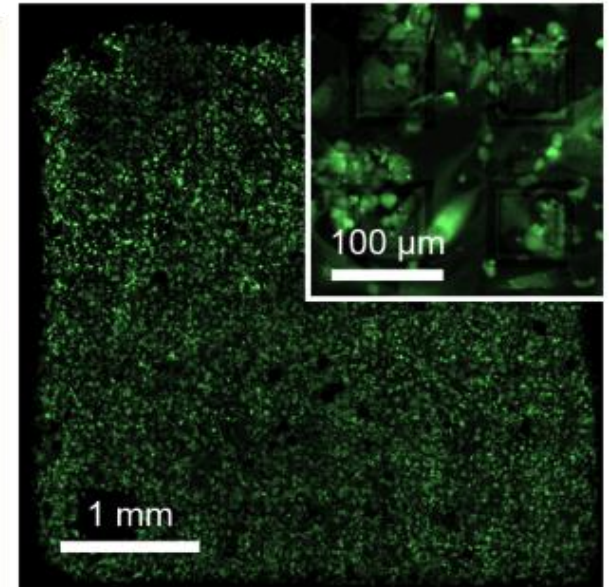
# Vertical nanoelectrodes on the surface + packaging



9 nanoelectrodes  
per pixel

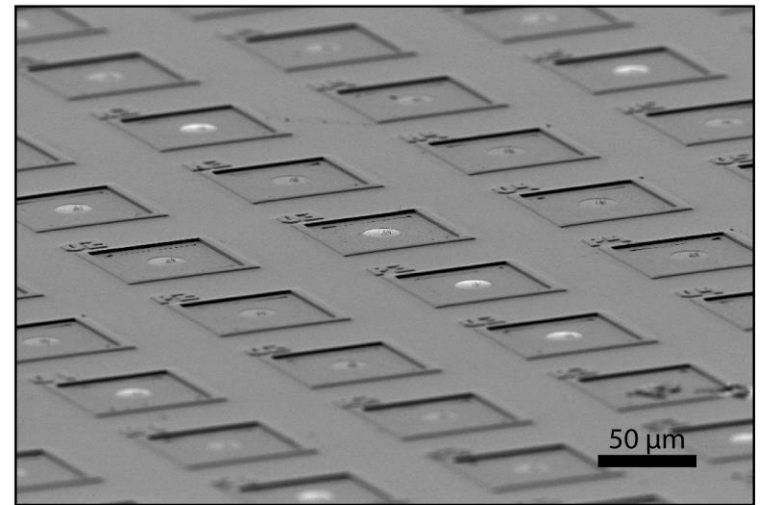
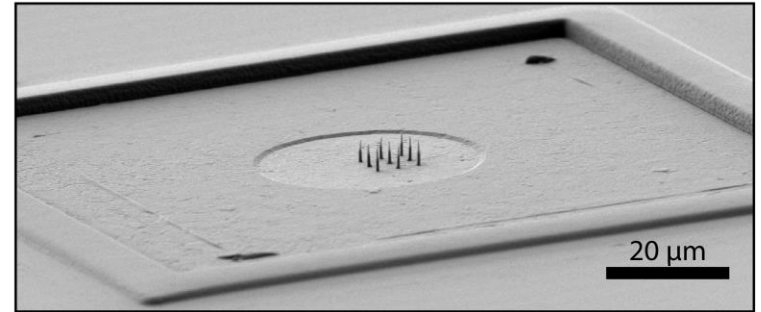
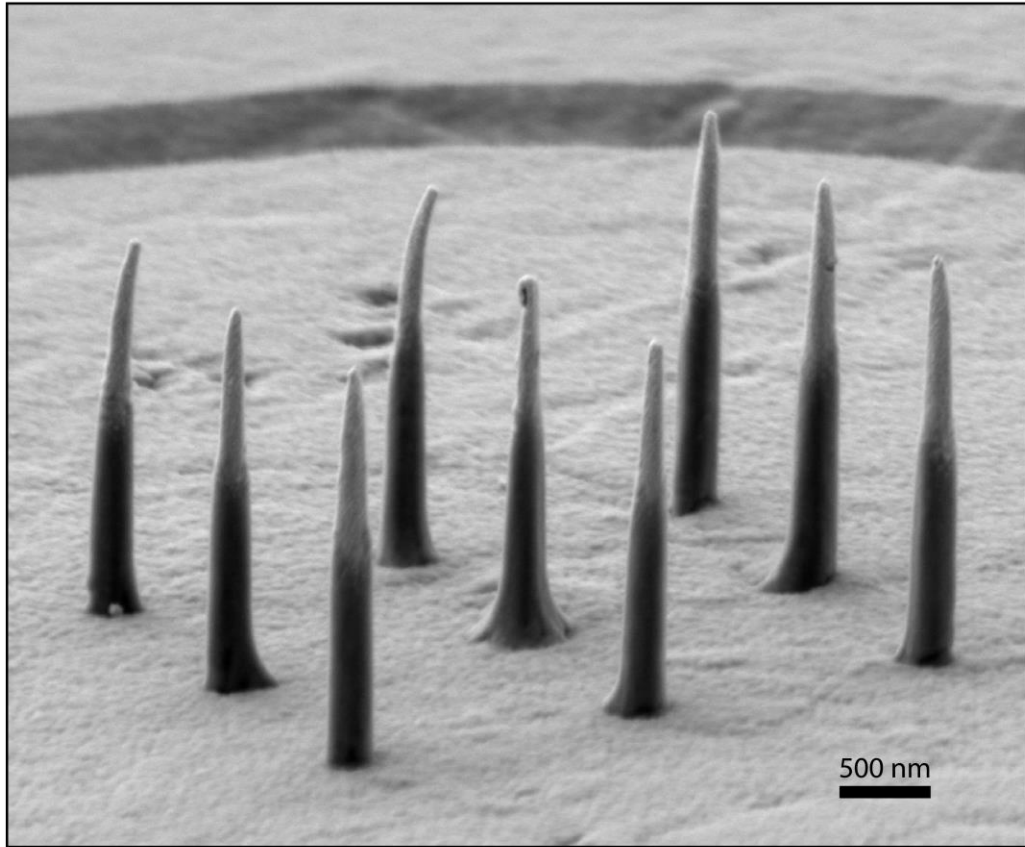


Packaged device



Cardiomyocyte tissue  
*in vitro* cultured on top

# Vertical nanoelectrodes



# Post fabrication steps



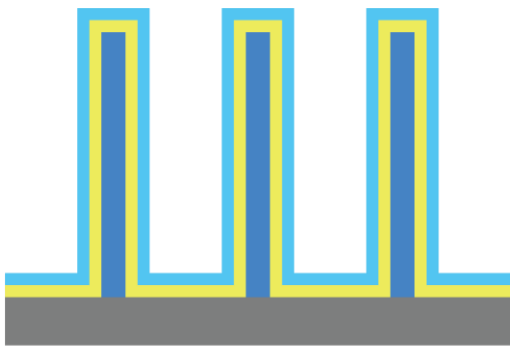
PECVD SiO<sub>2</sub> on Al pad



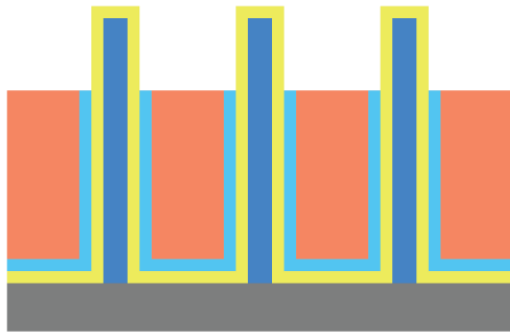
Stepper lithography and dry etch to form SiO<sub>2</sub> pillar



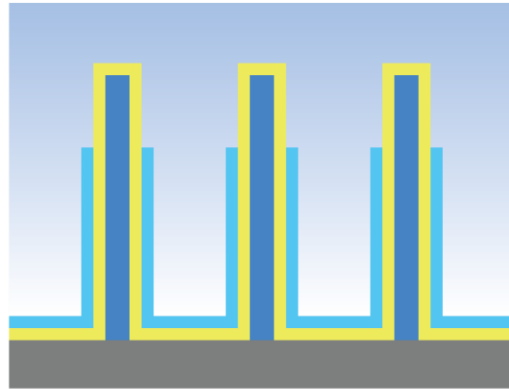
Wet etch pillar into SiO<sub>2</sub> wires



Sputter metal coating; insulate with ALD SiO<sub>2</sub>

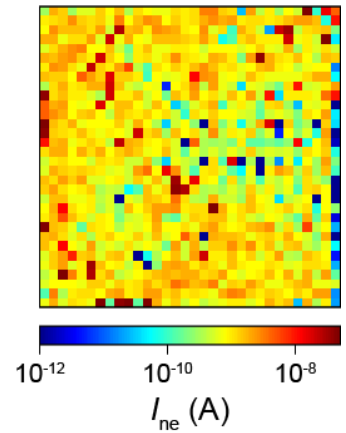
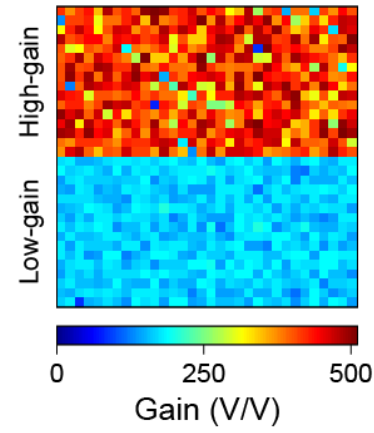
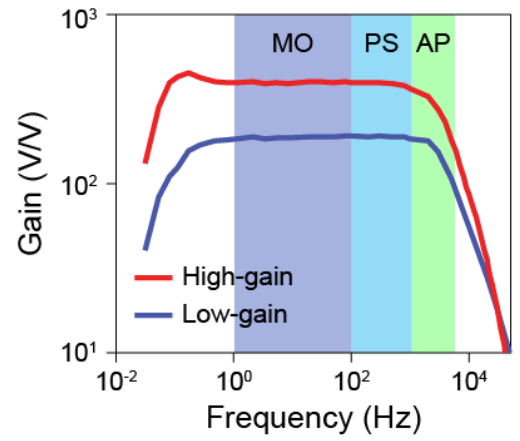
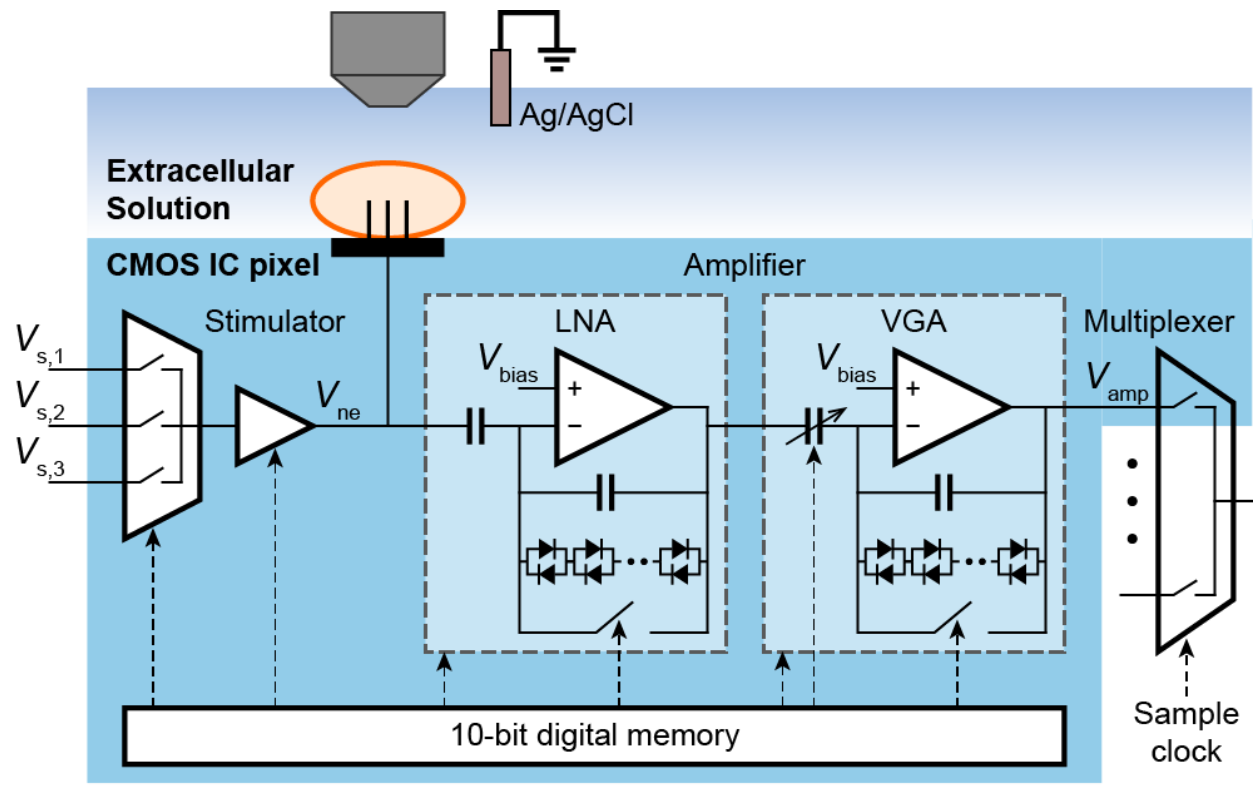


Spincoat resist; wet etch nanowire tip

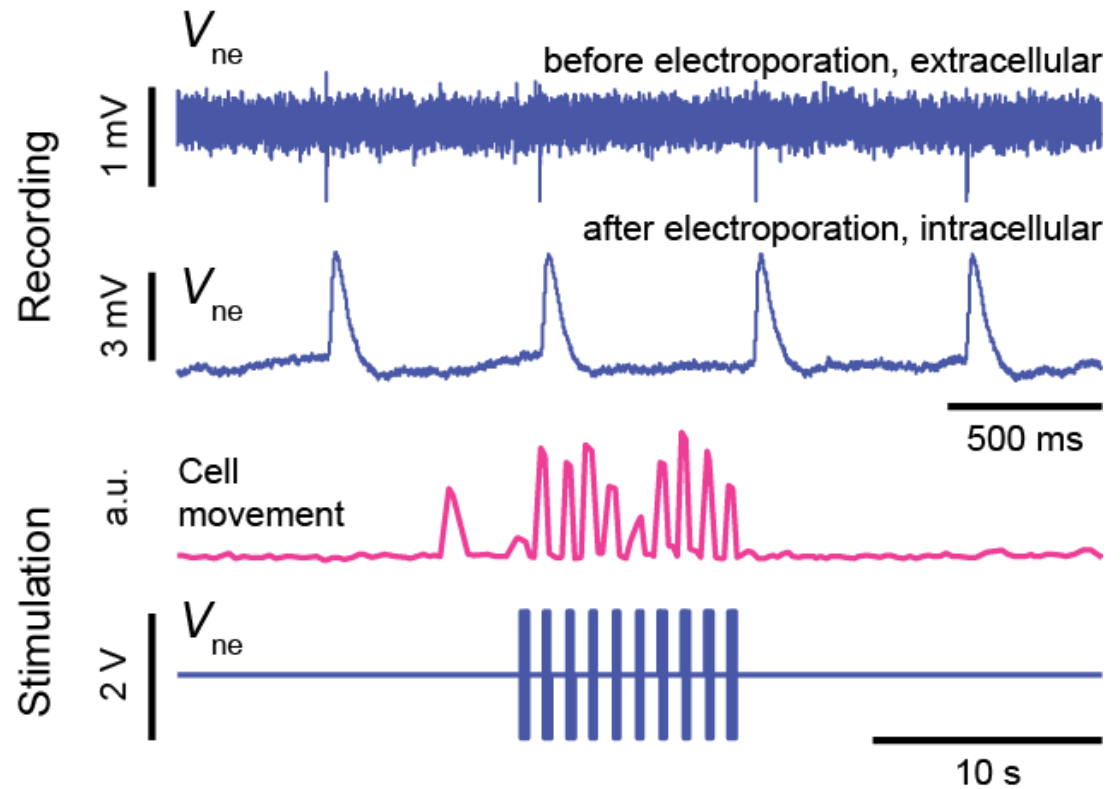
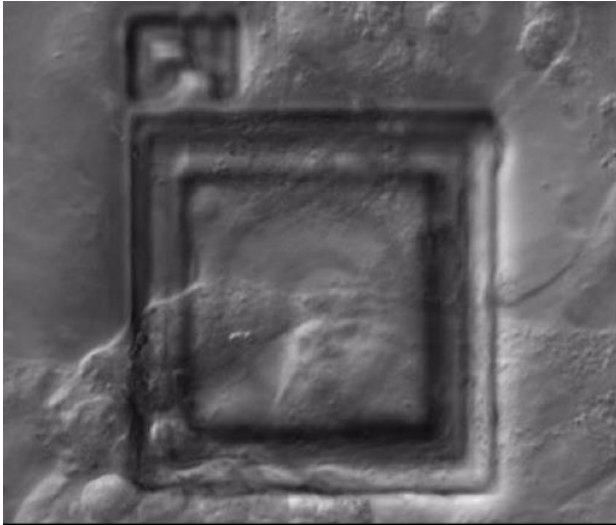


Ready for cell experiment in solution

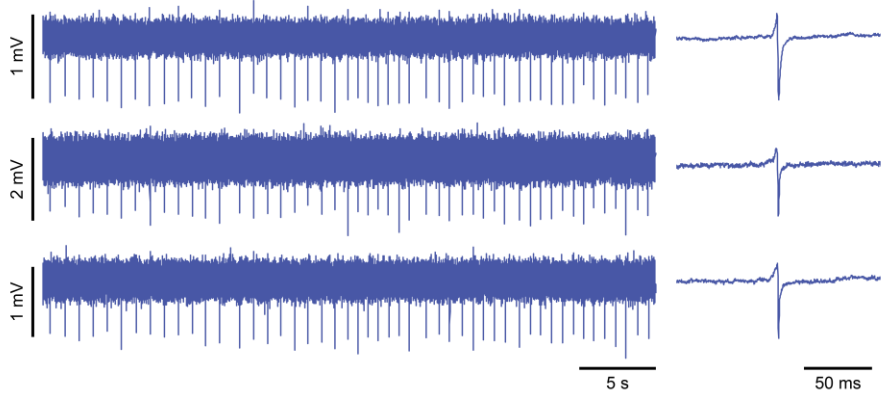
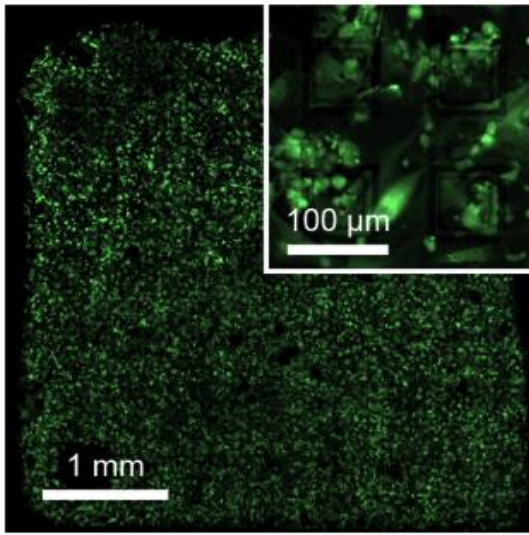
# Pixel circuit & electrode characterization



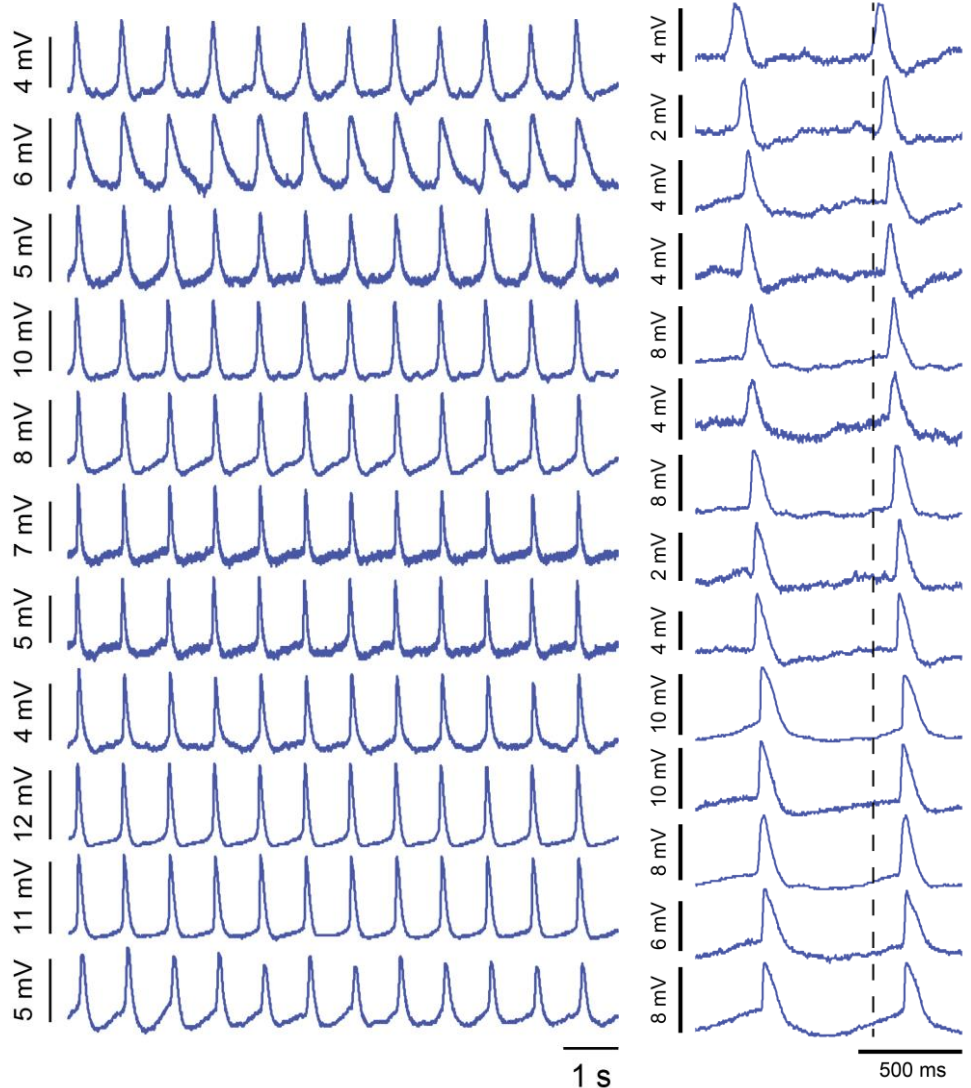
# Single myocyte intracellular recording & stimulation



# Parallel + intracellular recording from 235 cardiomyocytes

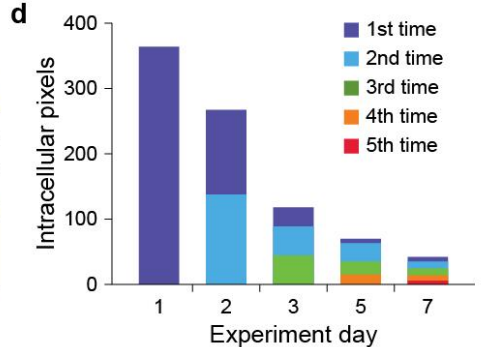
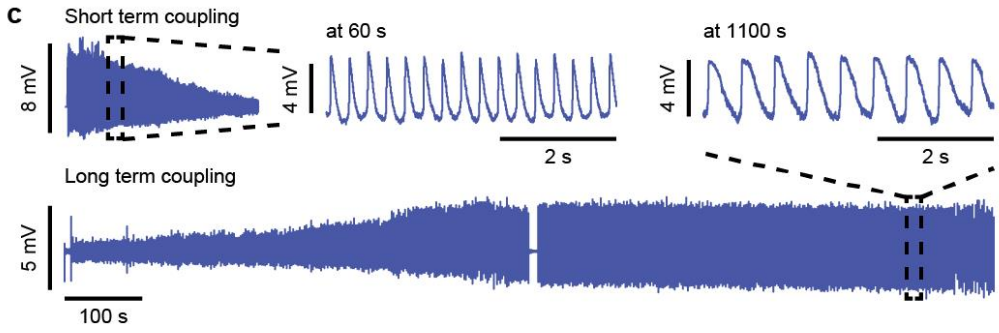
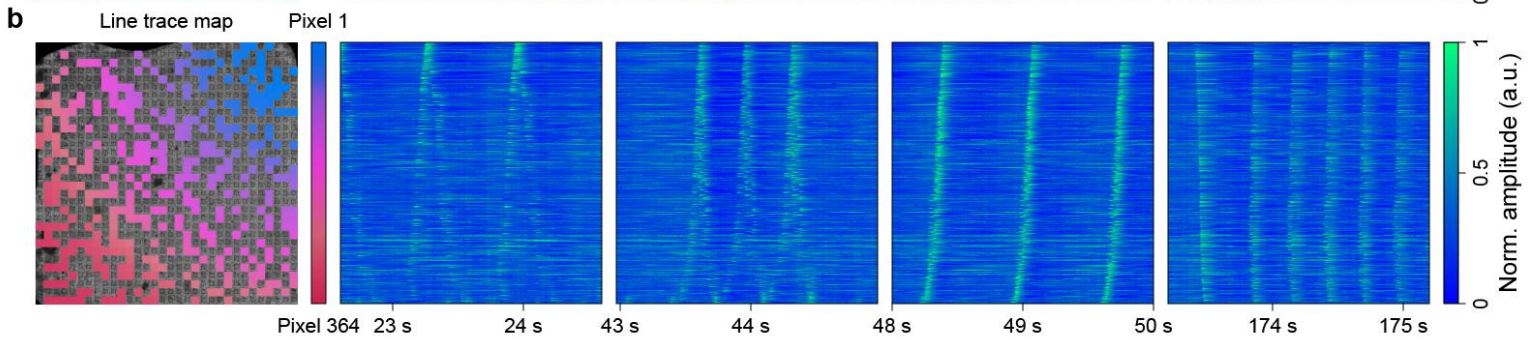
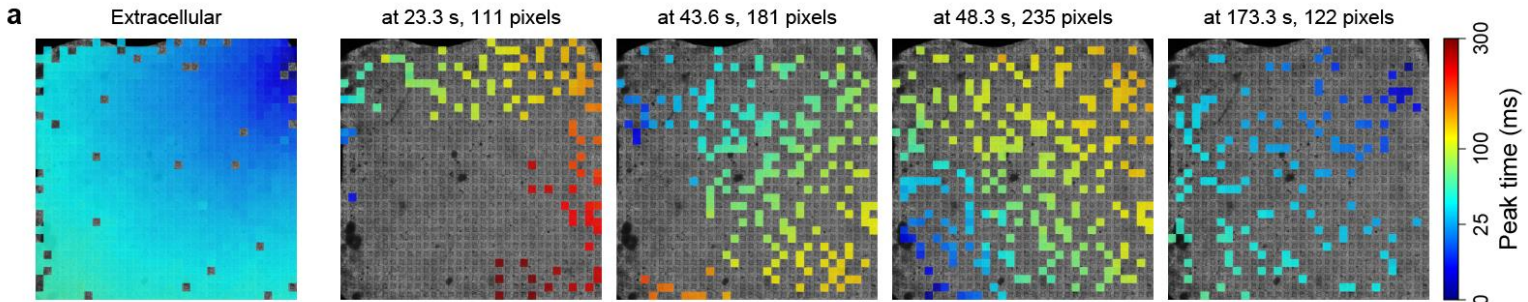


Extracellular recording

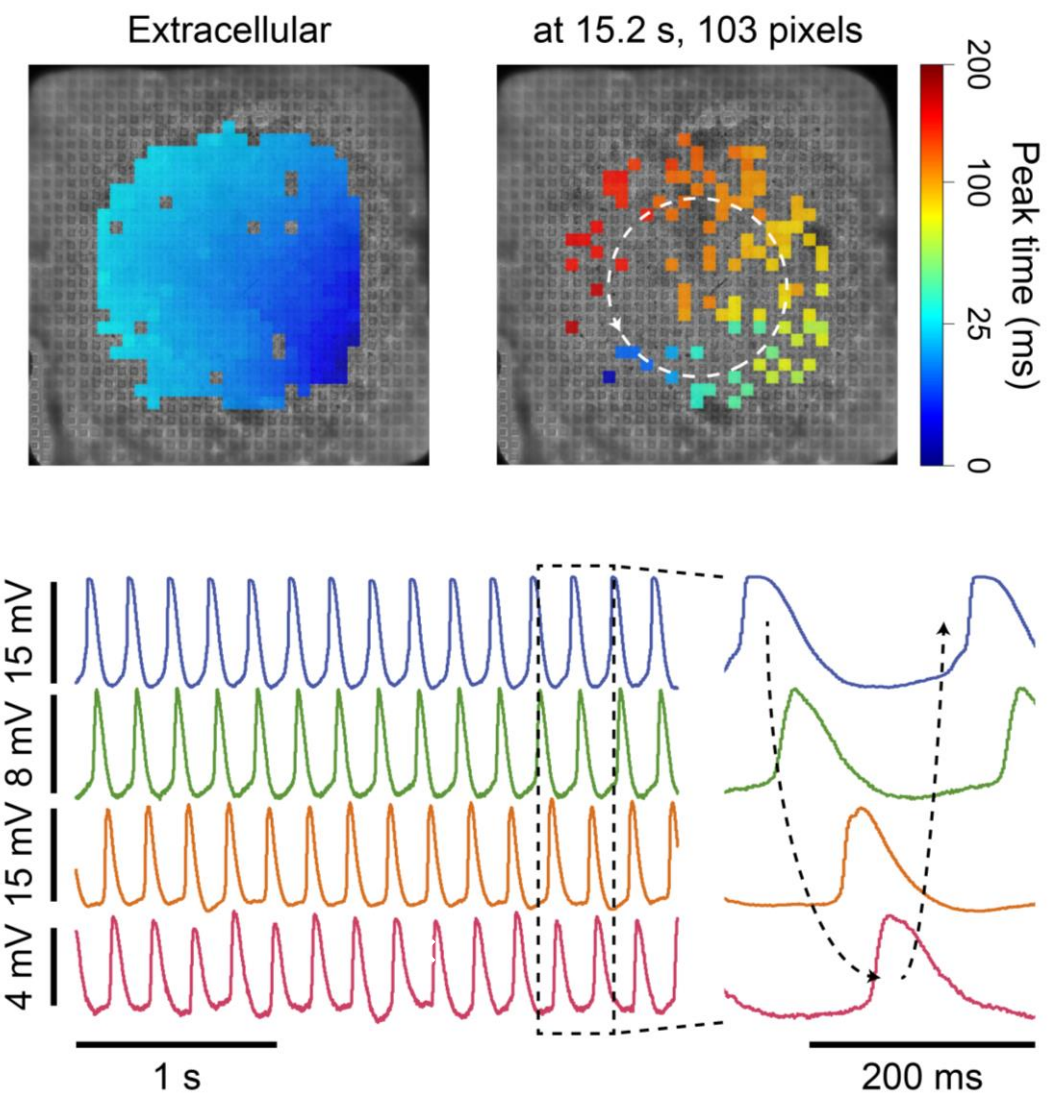


Intracellular recording

# Parallel + intracellular recording from 235 cardiomyocytes

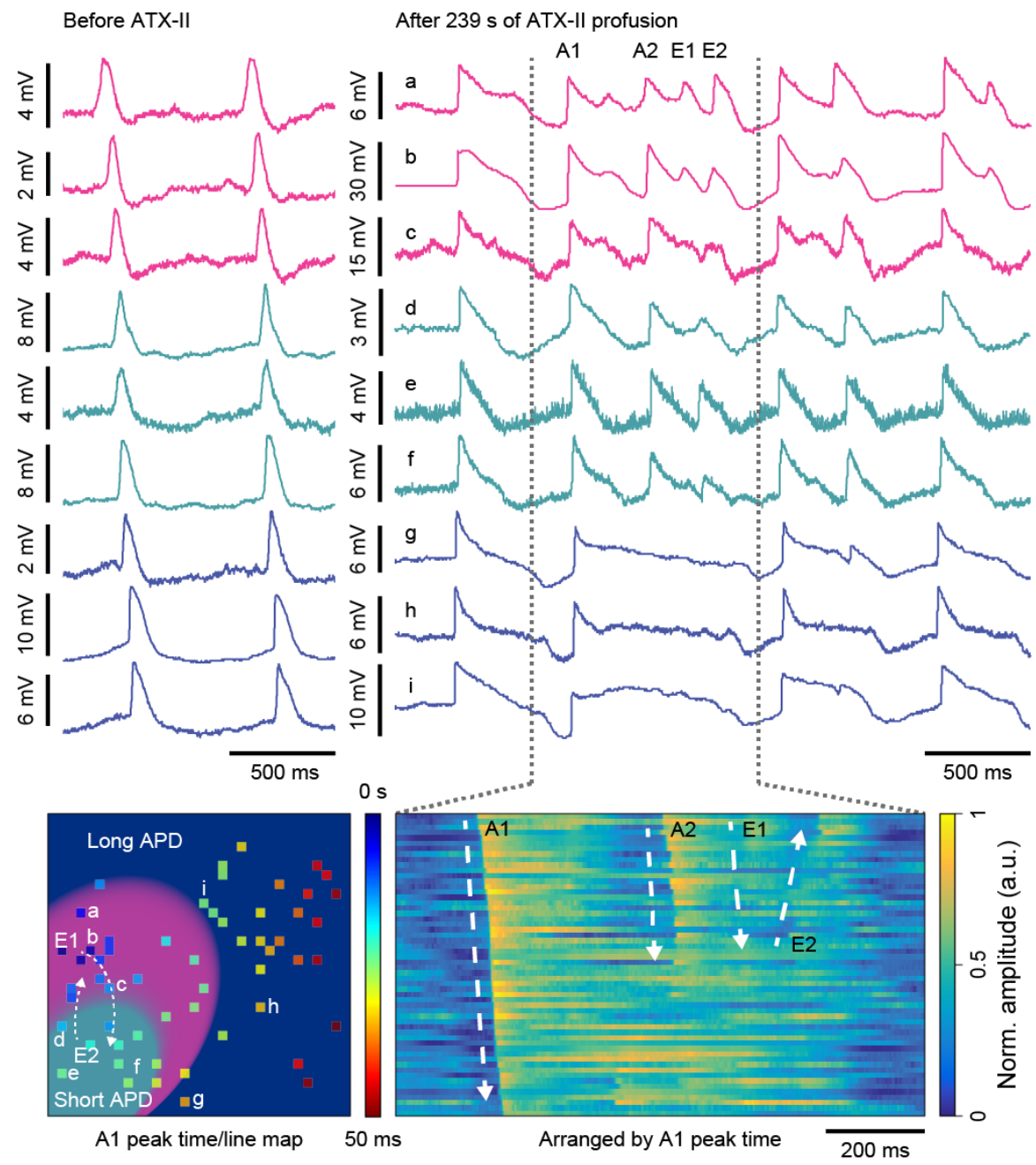


# Parallel + intracellular recording – another example





# Drug-screening — Network-level intracellular investigation



# CMOS nanoelectrode array for all-electrical intracellular electrophysiological imaging

Jeffrey Abbott<sup>1‡</sup>, Tianyang Ye<sup>1‡</sup>, Ling Qin<sup>1</sup>, Marsela Jorgolli<sup>2‡</sup>, Rona S. Gertner<sup>3</sup>, Donhee Ham<sup>1\*</sup> and Hongkun Park<sup>2,3,4\*</sup>

<sup>1</sup>School of Engineering and Applied Sciences, Harvard University, Cambridge, Massachusetts 02138, USA. <sup>2</sup>Department of Physics, Harvard University, Cambridge, Massachusetts 02138, USA. <sup>3</sup>Department of Chemistry and Chemical Biology, Harvard University, Cambridge, Massachusetts 02138, USA. <sup>4</sup>Broad Institute of MIT and Harvard, 415 Main Street, Cambridge, Massachusetts 02142, USA. †Present address: Hybrid Modality Engineering R&D, Amgen Inc., 1 Amgen Center Drive, Thousand Oaks, California 91360, USA. ‡These authors contributed equally to this work. \*e-mail: [donhee@seas.harvard.edu](mailto:donhee@seas.harvard.edu); [Hongkun\\_Park@harvard.edu](mailto:Hongkun_Park@harvard.edu)

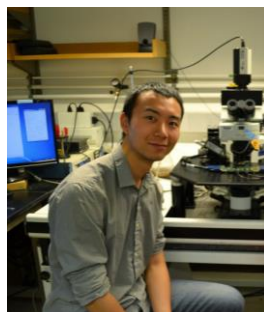


**ARL**

Catalyst Foundation



Jeffrey  
Abbott



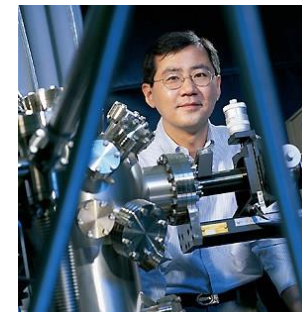
Tianyang Ye  
(Park)



Ling Qin



Marsela Jorgolli  
(Park)



Prof. Hongkun  
Park